

EV334000351US

BODY FLUID SAMPLER

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I. CROSS-REFERENCE TO RELATED APPLICATION

The present application is a continuation-in-part of Ser. No. 08/525,390, entitled "Interstitial Fluid Sampler," filed September 8, 1995 in the name of inventor 10 Paul D. Brinda and of U.S. Patent Application Ser. No. 08/525,942 entitled "Enhanced Interstitial Fluid Collection" and filed September 8, 1995 in the names of inventors Scott T. Latterell, Paul D. Brinda, Michael E. Hilgers, Michael J. Shoup, Thomas B. Hoegh and Brian J. 15 Erickson. The present application also discloses subject matter disclosed in commonly assigned U.S. Patent Application No. \_\_\_\_\_, entitled "Needle Assembly For Fluid Sampler" and filed concurrently herewith in the name of inventors Joel R. Racchini and Larry A. Walters.

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II. BACKGROUND OF THE INVENTION

1. Field of the Invention

This invention pertains to an apparatus for testing body fluid constituents. More particularly, this 25 invention pertains to a sampler for use in collecting body fluids such as interstitial fluid.

2. Description of the Prior Art

In the prior art, there are numerous examples of 30 apparatus for testing and determining the level of

constituents in human blood. A great deal of attention has been directed to the development of techniques and apparatus for measuring blood glucose.

As noted in commonly assigned and co-pending U.S.

5 Patent Application Nos. 08/321,305 and 08/136,304  
(corresponding to PCT International Publication No.  
WO95/10223 published April 20, 1995 on International  
Application No. PCT/US94/11580 and incorporated herein by  
reference), the determination of the level of a constituent  
10 of blood can be achieved by measuring the level of that  
constituent in other body fluids such as interstitial  
fluid. The aforementioned patent applications and  
international publication disclose a method and apparatus  
for a minimally invasive technique for collecting a sample  
15 of interstitial fluid through use of an extremely small  
needle which penetrates into the dermal layer of the skin  
in order to collect a low blood or blood-free sample of  
interstitial fluid. The collected interstitial fluid can  
then be analyzed for a determination of the level of  
20 constituents within the fluid. For example, the collected  
interstitial fluid can be analyzed for an amount of glucose  
with the determined amount being representative of the  
amount of glucose contained within the patient's blood.

The aforementioned applications and international  
25 publication disclose the use of a ring (item 60 in Fig. 6  
of the application) which surrounds the needle to create a  
pressure area on the patient's skin. It is believed this  
leads to an increase in the amount of interstitial fluid  
being collected.

In the collection of interstitial fluid, it is desirable to increase the speed at which a sample is collected. In the absence of mechanical or other assistance, the rate at which interstitial fluid is

5 collected through a small diameter tube or needle is very slow. Preferably, patients utilizing such equipment for home use will be provided with a system which collects interstitial fluid at a rapid pace to ensure that a patient does not remove the needle too early in its application.

10 Also, it is important to provide for techniques to increase a volume of interstitial fluid being collected through a needle.

When collecting any body fluid through use of a needle, it is important that the needle be a disposable item in order to prevent re-use of the needle. Such re-use can result in the transmission of disease. Where the apparatus is to be used in a patient's home by the patient, the apparatus should be simple to use and with the needle incorporated in a disposable item. Since the needle is

20 incorporated in a disposable item, it is important that the disposable item be amenable to low-cost manufacture. Also, in order to test the interstitial fluid, the interstitial fluid collection mechanism must be coupled with an analytic mechanism for analyzing the collected fluid. Where such a

25 device is to be used in home by low-skilled patients, it is important that the sampler and the analytic portion of the device be mutually configured to ensure that the sampler is coupled to the apparatus in a repeatable and reliable

manner to minimize errors resulting from use of the apparatus by untrained patients.

### III. SUMMARY OF THE INVENTION

5 According to a preferred embodiment of the present invention, a sampler is disclosed for use in a body fluid collection apparatus where the collection apparatus has a light source for generating a testing light and a light detector for detecting light. The light source and  
10 the detector are contained within the apparatus in a predetermined alignment to define a light path between the source and the detector. The apparatus further includes an opening of predetermined geometry to define an access to the light path. The sampler includes a main body having a  
15 handle and a sample end. The sample end has an external mating geometry which mates with the predetermined geometry of the opening of the apparatus. Therefore, the sample end can be inserted into the opening in a predetermined alignment such that the sampler may be repeatably inserted  
20 within the opening in the predetermined alignment with a sample location of the sample end positioned within the light path. A body fluid collection apparatus is carried on the sampler. The body fluid collection apparatus includes a needle sized to protrude beyond the housing.  
25 The needle protrudes a distance selected for the needle to penetrate into a body fluid-laden skin layer when the housing is urged against the skin layer by the user. An absorbent medium is carried on the sampler in fluid flow communication with the needle for body fluid to flow from

the needle onto the medium. The medium is positioned at the sample location.

IV. BRIEF DESCRIPTION OF THE DRAWINGS

5 Fig. 1 is a cross-sectional elevation view of an interstitial fluid apparatus showing a sampler contained within the apparatus in a retracted position;

Fig. 2 is the view of Fig. 1 with the apparatus shown in an extended position;

10 Fig. 3 is a perspective exploded view of the apparatus of Fig. 1;

Fig. 4 is the view of Fig. 3 rotated 90° to the right of the view of Fig. 3;

15 Fig. 5 is the view of Fig. 4 rotated 90° to the right of Fig. 4;

Fig. 6 is the view of Fig. 5 rotated 90° to the right of Fig. 5;

Fig. 7 is a perspective view of an optics housing for use in the apparatus of Fig. 1;

20 Fig. 8 is a side elevation view of the housing of Fig. 7;

Fig. 8A is an enlarged view of a bottom portion of the view of Fig. 8;

25 Fig. 9 is a side elevation view of the housing of Fig. 7 rotated 90° from the view of Fig. 8;

Fig. 10 is a view taken along line 10-10 of Fig. 9;

Fig. 11 is a bottom plan view of the housing of Fig. 7;

Fig. 12 is a perspective view of a shell for use  
in the apparatus of Fig. 1;

Fig. 13 is a sectional view of the shell of Fig.  
12;

5 Fig. 14 is a perspective view of a collar for use  
in the apparatus of Fig. 1;

Fig. 15 is a sectional view of the collar of Fig.  
14;

10 Fig. 16 is a perspective view of a base for use  
in the apparatus of Fig. 1;

Fig. 17 is a sectional view of the base of Fig.  
16;

15 Fig. 18 is a top, left side and rear end exploded  
perspective view of a sampler for use in the apparatus of  
Fig. 1;

Fig. 19 is a top, left side and rear end  
perspective view of a sampler main body for the sampler of  
Fig. 18;

20 Fig. 20 is a left side elevation view of the  
sampler main body of Fig. 18 (with the opposite side being  
substantially identical);

Fig. 21 is a view taken along line 21-21 of Fig.  
20;

25 Fig. 22 is a view taken along line 22-22 of Fig.  
20;

Fig. 23 is an enlarged bottom view of a front  
portion of the main body of Fig. 20;

Fig. 24 is a side elevation view of a piston for  
the sampler of Fig. 18;

Fig. 25 is a view taken along line 25-25 in Fig.  
24;

Fig. 26 is a side elevation view of a catch pin  
for the sampler of Fig. 18;

5 Fig. 27 is a view taken along line 27-27 in Fig.  
26;

Fig. 28 is a schematic representation showing the  
apparatus of Fig. 1 placed against a patient's skin;

10 Fig. 29 is the view of Fig. 28 showing initial  
forcing of the apparatus against the patient's skin;

Fig. 30 is the view of Fig. 28 showing urging of  
the apparatus against the patient's skin with penetration  
of a needle into the patient's skin layer and with a  
piston aligned with a pressure ring;

15 Fig. 31 is the view of Fig. 28 with the piston  
protruding beyond the pressure ring;

Fig. 32 is a top, front, right side perspective  
view of an alternative embodiment of the present invention;

20 Fig. 33 is an exploded view of the alternative  
embodiment of Fig. 32;

Fig. 34 is a cross-sectional view of the  
embodiment of Fig. 32;

Fig. 35 is a transverse cross-sectional view of  
the embodiment of Fig. 32;

25 Fig. 36 is a partial sectional view of the  
embodiment of Fig. 32 showing initial insertion of a  
sampler into the apparatus;

Fig. 37 is the view of Fig. 36 showing further insertion of the sampler into the apparatus for a cam to engage a cam follower;

5 Fig. 38 is the view of Fig. 37 showing still further insertion of the sampler with a sample end pivoted to a sample position and with the sampler not yet fully inserted;

Fig. 39 is the view of Fig. 38 with the sampler fully inserted;

10 Fig. 40 is the view of Fig. 39 with the sampler partially withdrawn and with the sample end partially pivoted to a storage position by reason of the cam acting against a second cam follower;

15 Fig. 41 is an enlarged view of relative positioning of a needle on a sampler with a membrane not shown;

Fig. 42 is a perspective view of a needle and membrane assembly; and

Fig. 43 is a cross-sectional view of Fig. 42.

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#### V. DESCRIPTION OF THE PREFERRED EMBODIMENT

With reference now to the various drawing figures in which identical elements are numbered identically throughout, a description of the preferred embodiment of the present invention will be shown. While the invention will be described with reference to an apparatus for collecting interstitial fluid to test for glucose within the interstitial fluid, it will be appreciated that the apparatus can be used for testing any

body constituent which may be contained within interstitial fluid.

In a preferred embodiment, the apparatus is disclosed with reference to use of a penetrating needle and

5 an absorbing membrane such as that shown and described in U.S. Patent Application Serial Nos. 08/321,305 and 08/136,304 (and corresponding PCT International Publication No. WP 95/10223, dated April 20, 1995 on International Application No. PCT/US94/11580, incorporated herein by

10 reference). With reference to Figs. 16-20 of that application (showing a representative embodiment of the invention shown in that application), a needle 214' is surrounded and maintained in fixed relative position by a ring 202'. The ring is placed against a patient's skin in

15 order to define a pressurized area on the patient's skin as the needle 214' penetrates into the skin. The needle is sized to be about 28 to 32 gauge (i.e., .36 mm outside diameter to .23 mm outside diameter) with an anticipated preferred size of about 30 gauge. The needle is made as

20 small as possible to provide a minimally intrusive and painless insertion into the skin. The needle is sized to penetrate into the dermis for a variety of reasons as best disclosed in the aforementioned application including low pain and the collection of low blood interstitial fluid for

25 subsequent testing. An absorbent membrane 210' is placed in fluid flow communication with the needle 214' such that interstitial fluid which flows through the needle 214' is deposited on the membrane 210' as a spot available for subsequent testing with light (visible or non-visible

spectrum). The amount of absorption of various wavelengths of the light indicates the concentration of constituents for testing such as glucose or the like.

The present invention pertains to a testing apparatus which includes a needle 10 disposed in fluid flow communication with an absorbent membrane 12 both in accordance with the teachings of the aforementioned PCT International Publication No. W095/10223.

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#### A. First Described Embodiment

The present invention is directed to an apparatus 1-6) for collecting and testing interstitial fluid. The apparatus 20 includes a main housing 22 (shown in Figs. 1 and 2 only) coupled to a base 24. The apparatus 20 further 15 includes a collar 26 secured to the base 24. A shell 28 is contained within the collar 26. An optics housing 30 is contained within the shell 28. Finally, a sampler 32 is provided to be received within the optics housing 30. Each of base 24, collar 26, shell 28, optics housing 30 and 20 sampler 32 will be separately described.

Main housing 22 is shown only in section in Figs. 1 and 2. Main housing 22 is sized to be gripped by a patient such that the apparatus 20 may be urged against the patient's skin for purpose of collecting interstitial fluid 25 as will be described. In addition to constituting a handle which can be grasped by the patient, the main housing 22 will contain electronics and the like for generating power for a light source as will be described and for analyzing signals from a light detector (as will be described) in

order to calculate the level of constituents, such as blood glucose, contained within a sample of interstitial fluid.

Such electronics are not shown but it will be appreciated that such electronics are well within the skill of the art.

- 5 Examples of circuits for analyzing sampling light are described in commonly assigned U.S. Patent No. 5,115,133 to Knudson dated May 19, 1992 and the aforementioned International Publication No. W095/10223.

The base 24 is separately shown in Figs. 16 and  
10 17. Base 24 is substantially cylindrical and is provided with an end plate 36 having holes 38 extending at least partially therethrough with the holes 38 sized to receive any suitable fastening means such as bolts or the like for fastening of the end plate 36 to the main housing 22. The  
15 base 24 further includes an inner hollow cylinder 40 extending from plate 36 with the inner cylinder 40 being coaxial with an outer cylinder 42 of the base 24. Outer cylinder 42 has a threaded inner surface 44.

The collar 26 is separately shown in Figs. 14 and  
20 15. The collar 26 includes an enlarged cylindrical portion 50 sized to be received within base 24 and with an end 51 abutting the end plate 36 of base 24. An outer wall 52 is threaded to mate with the internal threading 44 of base 24. An inner wall 53 of cylindrical portion 50 remains spaced  
25 from inner cylinder 40 to define a void for receiving springs as will be described (and as shown in Figs. 1-2). The collar 26 also includes a reduced diameter portion 54 with the reduced diameter portion 54 and the enlarged diameter portion 50 connected at an annular stop surface 56

shown in Fig. 15. For purposes that will become apparent, the reduced diameter portion 54 includes a slot 58 at an end 59 of portion 54. Linearly aligned with slot 58 is a hole 61.

5           The shell 28 is separately shown in Figs. 12 and 13. The shell 28 includes a cylindrical body 60 sized to be slideably received in close tolerance within the reduced diameter cylindrical portion 54 of collar 26. The cylindrical body 60 terminates at a flange 62 positioned to 10 abut stop surface 56 of collar 26. Accordingly, the shell 28 is slideable within the collar 26 with the flange 62 movable between the stop surface 56 of collar 26 and the end plate 36 of base 24.

The cylindrical body 60 has at its end opposite 15 flange 62 a reduced diameter portion 64 which is coaxial with the main cylindrical body 60. The reduced diameter portion 64 terminates at a first pressure ring 66 with the plane of the opening of the pressure ring 66 being generally perpendicular to the cylindrical axis of body 60. 20 An elongated slot 68 extending generally in the direction of the axis of body 60 is provided extending through the shell 28 with the slot 68 extending substantially the length of the body 60 and substantially the length but not entirely through the sidewall of the reduced diameter 25 portion 64 such that ring 66 is an uninterrupted ring. However, a segmented ring or other incomplete ring would be satisfactory.

The optics housing 30 is separately shown in Figs. 7-11 and includes a generally cylindrical main body

70 (with flat side walls 71a, 71b) having extending axially therefrom a reduced diameter cylinder 72 (surrounded by surface 71) having an annular slot 73. The reduced diameter cylinder 72 is sized to be slideably received  
5 within the inner cylinder 40 of base 24 as best shown in Figs. 1 and 2.

The main body 70 includes a first axial slot 74 extending partially through a distal end 75 of the body 70. Disposed axially spaced from slot 74 is a second slot 76  
10 extending through the main body 70. A pin receiving hole 77 extends through body 70 perpendicular to slot 76. Ninety degrees offset from slots 74, 76 are access holes 78 in communication with a hollow interior 80 of cylinder 72. Ninety degrees offset from slot 74 are pockets 82, 83 with  
15 axes of the pockets 82, 83 in coaxial alignment with one another and in communication with the slot 74. The base end 75 has a ramped ridge 79 extending parallel to hole 77.

In the assembly, as best shown in Figs. 1 and 2, a first biasing spring 84 is positioned to act between the  
20 base plate 36 of base 24 and the flange 62 of shell 28 urging the shell 28 away from the base plate 36. A second biasing spring 86 is positioned to act against the base plate 36 of base 24 and an engaging surface 71 on cylinder 70 thereby urging the optics housing 30 axially away from  
25 the base plate 36.

As shown in Figs. 3-6, a light source 90 is contained within pocket 82. A light detector 92 is contained within pocket 83. Electrical leads (not shown) from both the light source 90 and light detector 92 may be

passed between the opposing exterior surfaces 71a, 71b of cylinder 70 and the interior surface of shell cylinder 60 with the leads then passed through the holes 78, into hollow interior 80 of cylinder 72 and directed thus into 5 the circuitry (not shown) contained within the housing 22. The light source 90 and light detector 92 are aligned to define a light path therebetween. The light source 90 generates a testing wavelength. The light detector 92 is selected to measure the intensity of wavelengths including 10 the intensity of the testing wavelength.

A lock pin 94 (shown separately in Figs. 26-27) is contained within optics housing 30 in hole 77 with the lock pin 94 positioned at a 90° angle to the plane of the slot 74. The pin 94 has a ramp 95 disposed in slot 76. In 15 the assembly shown in Figs. 1-6, the slots 74, 76 of the optics housing 30 are in alignment with the slot 68 of the shell 28.

As shown in Figs. 18-25, the sampler 32 includes a body 100 formed of injection molded plastic. The body 20 100 includes a rear handle portion 101 and a forward sampling portion 102. The handle portion 101 is sized to be gripped by the fingers of a user. At the sampling end 102, the body 100 is provided with a hub or piston 104. The piston 104 is cylindrical and sized to be received in 25 close sliding tolerance within the reduced diameter cylinder 64 of shell 28. The piston terminates at a flat second pressure surface 106 which is generally perpendicular to the axis of the needle 10. While a flat

surface 106 is preferred, other shapes (e.g., concave) could be used.

The needle 10 protrudes beyond the surface 106 a distance equal to a desired penetration of the needle 10 into a patient's skin layer. As disclosed in the aforementioned international publication, distance of protrusion of needle 10 is about 1.5 mm to ensure protrusion of the needle 10 into but not through a dermal layer of a patient's skin. At the sampling end 102, the main body 100 is provided with a relief 108 surrounding a hole 110 formed through the body. The hole 110 is in communication with a proximal end 11 of the needle 10. Accordingly, an absorbent material 12 such as the material 210' shown in Figs. 16-20 of the aforementioned International Publication No. W095/10223 may be placed within the relief 108 such that interstitial fluid which flows up the needle 10 will be deposited upon the material 12. The material 12 is held in place through any suitable means such as by an adhesive ring 111 (or, alternatively, ultrasonic bonding or other bonding technique).

The hole 110 is positioned at a sampling location such that the hole 110 is in the light path between the light source 90 and the light detector 92 when the sampler 32 is placed within the apparatus 20 as will be described. The end 102 is sized to be received within the aligned slots 68, 74 of shell 28 and optics housing 30, respectively.

The main body 100 is provided with an arcuate rib 113 sized and shaped to abut an exterior surface of

the optics housing 30 on both sides of the slot 74 and to curve beneath the base 75. A latching member 112 is connected to the body 100. The latching member 112 pivots at a point of connection to the body 100 and includes a lever arm 114 exposed at the handle portion 101 such that the lever member 114 may be depressed manually by a user. The latch 112 further includes a latching end 116 sized and positioned to be received within the hole 76 of the optics housing 30. The latching end 116 includes a detent 118 (Figs. 1-2) positioned to engage and receive the ramp 95 of the lock pin 94 within the detent 118 when the sampler 32 is inserted within the slots 74, 76 in a predetermined alignment and with the sampling location 110 disposed within the light path between the source 90 and detector 92. A leading end of the locking end 116 is provided with a ramped surface to ride over the pin 94 upon insertion of the sampler 32 within the optics housing 30 and to provide a positive lock as the pin is received within the detent 118. To further secure the sampler 32 in optics housing 30 in the desired alignment, sampler housing 100 has a detent 117 (Fig. 23) to receive ridge 79 on the base 75 of optics housing 30. The sampler 32 may be easily removed by a user depressing end 114 thereby raising end 116 for the pin 94 to clear the detent 118 permitting removal of the sampler 32 from the apparatus.

With the construction thus described, a sampling end 102 may be placed within the aligned slots 74, 68. Over-insertion is avoided by reason of the sampling end 102 butting up against the interior of the optics housing 30.

Further, the lock pin 94 received within the detent 118 and the ridge 79 in detent 117 ensure that the sampler 32 is not under-inserted into the slots 74, 76 by providing a user with a positive feedback indicating that the lock pin

5 94 has been received within the detent 118 indicating the sampler 32 is in the predetermined alignment. Accordingly, upon receipt of such feedback, the user is assured that the sampling location 110 is in alignment with the light path between the light source 90 and the light detector 92.

10 The first spring 84 urges the shell away from the base 24 such that the full length of the piston 104 and needle 10 may clear the first pressure ring 66 and be inserted through the slot 68 as the sampler 32 is loaded into apparatus 20.

15 Due to the locking at detents 118 and 117, sampler 32 is held in a predetermined alignment with the membrane 12 in the light path between light source 90 and light detector 92. To facilitate placement of sampler 32 within apparatus 20, the sampler 32 and apparatus 20 have  
20 mating external geometries. Namely, in the rest position of Fig. 1, the shell 28 is fully extended from base 36 by spring 86. Slot 58 of collar 26, slot 68 of shell 28 and slot 74 of optics housing 30 are aligned to permit insertion of end 102 of sampler 32. Further, in this  
25 position, slot 68 is sized so that needle 10 may pass ring 66 without interference. Also, in this position, slot 61 of collar 26, slot 68 of shell 28 and hole 76 of optics housing 30 are aligned to receive end 116 of lever arm 112.

Upon insertion, the mating geometry of sampler 32 and optics housing 30 insure the membrane 12 is accurately positioned. The ribs 113 acting against the external surface of optics housing 30 together with ribs 95, 79 5 received within detents 118, 117 securely couple the sampler 32 to optics housing 30 in accurate alignment and with the sampler 32 movable with the optics housing 30. As the optics housing 30 moves relative to shell 28 and collar 26, the sizing of slots 58, 61 and 68 avoid interference 10 with movement of the sampler 32.

Upon initial placement of the apparatus against a patient's skin 200 (Fig. 28), the ring 66 first contacts a patient's skin 200 with the needle 10 being recessed behind the ring 66. Upon urging of the apparatus 20 against the 15 skin 200, the ring 66 moves relative to the needle 10 against the bias of the first spring 84. Upon achieving such relative movement, the needle 10 then penetrates the skin 200 with the second pressure surface 106 of the piston 104 engaging the skin and with both springs 84, 86 20 resisting further penetration until both springs are compressed. Second spring 86 ensures a constant force acts on piston 106.

Figs. 28-30 show a sequence of operation of the present apparatus. As shown in Fig. 28, during the rest 25 state, the needle 10 is recessed behind the first pressure ring 66 to prevent damage to the needle 10 and inadvertent skin penetration. Upon initial urging of the pressure ring 66 against the skin (Fig. 29), the pressure ring 66 depresses the skin 200 and makes the skin taut in the area

defined by the ring 66. Further, the pressurization creates a pressurized area in the zone of the skin layer 200 directly beneath the ring 66. This is desirable since interstitial fluid beneath the skin 200 is believed to

5 exist at a negative pressure. Creating a pressurized zone beneath the ring 66 is believed to assist in rapid collection of interstitial fluid within the needle 10.

During this initial pressurization of the skin 200, the ring 66 moves relative to piston 104 until the needle 10

10 penetrates the skin 200 and the end 106 of the piston 104 abuts the skin 200 (Fig. 30). Further depression (which can occur against soft skin but which may not occur against more rigid skin) is shown in Fig. 31 where the piston end surface 106 protrudes slightly beyond the ring 66 to

15 further increase the pressure acting in the collection zone of the skin 200 and with full penetration of the needle 10.

It has been found that this sequence of action significantly increases the rate at which interstitial fluid is collected through the needle 10 and deposited on

20 the membrane 12 within the sampler 32.

After full penetration of the needle 10, internal circuitry may then be actuated to operate the light source 92. Absorption of the testing light through the collected sample provides an indication of the amount of the

25 constituent contained on the sample.

In a preferred embodiment, springs 84, 86 are preloaded. Namely, in the rest position of Figs. 1 and 28, first spring 84 exerts an urging force on shell 28 of about three pounds and with a spring constant of about four

pounds per inch. Spring 86 is pre-loaded to about one pound and has a spring constant of about two pounds per inch. To accommodate the pre-loading of springs 84, 86, optics housing 30 is provided with a retaining ring 202 5 (shown only in Figs. 1 and 2) in slot 73. The pre-loading of spring 84 insures a minimum skin pressure by ring 66 before penetration of the skin 200 by needle 10.

As shown best in Figs. 1, 2 and 18, membrane 12 is provided with a U-shaped boundary 300. Boundary 300 is 10 formed by ultrasonically or pressure treating membrane 12 to create a material density in boundary 300 which is greater than a material density of the remainder of the membrane 12. Therefore, boundary 300 provides an increased resistance to liquid flow compared to the remainder of the 15 absorbent membrane 12. The end 11 of needle 10 is positioned to deposit interstitial fluid onto the interior of the U-shaped boundary 300. The increased density of the boundary 300 permits the fluid to flow within the interior of the boundary 300 but restricts fluid flow beyond the 20 boundary 300. The target location ("T") of light through membrane 12 during testing is positioned within the boundary 300. Boundary 300 thus insures that a sufficient volume of collected fluid is in residence at the target location T during testing.

25 It will be appreciated that through use of the present invention the rate at which interstitial fluid is collected through the needle 10 is greatly enhanced over that shown in the aforementioned International Publication No. WO95/10223. Further, the sampling apparatus is

contained within the low-cost sampler 32 which can be readily disposed of after each use. The mating geometry of the sampler 32 with the internal geometry of the apparatus 20 ensures that the sampler 32 is placed within the 5 apparatus 20 in a predetermined alignment with the sampling location in the light path between the source 90 and the detector 92. The sampling apparatus also ensures a proper positive locking position which may be released easily by an operator and the entire operation of insertion of the 10 sampler within the apparatus and removal of the sampler for subsequent disposal is easily accomplished for a patient..

B. Second Described Embodiment

Figs. 32 through 39 illustrate an alternative 15 embodiment of the present invention. In these figures, the base 24 of the previous described embodiment as well as collar 26, shell 28 and optics housing 30 are replaced with a piston housing 400 which slideably receives two hollow pistons 402. The pistons 402 are disposed to move in generally parallel paths of travel relative to the piston 20 housing 400.

The piston housing 400 is retained in a stationary position (relative to tool handle 22 of Fig. 1) by means of clam shell back 404 which is adjoined to clam 25 shell front 406. The clam shells 404, 406 may be connected to the apparatus housing 22 as in the previous described embodiment. The clam shell front 406 has an axially extending slot 408. Bottom ends of the hollow pistons 402 are secured to a pressure ring shell 412 by means of screws

or bolts 414 (Fig. 34). An optical core 420 is provided with the pistons 402 passing through aligned holes 422 of the core such that the core 420 is movable relative to the piston housing 400 and the pistons 402. A flange 403 on 5 clam shells 404, 406 limits downward movement (in the view of Fig. 34) of optical core 420.

Best shown in Fig. 34, outer springs 416 surrounds the pistons 402 between the piston housing 400 and the optical core 420 such that springs 416 compress 10 when the movable optical core 420 moves upwardly (in the view of Fig. 34) relative to the stationary piston housing 400. Further, springs 418 are positioned within each of the hollow pistons and opposing the housing 22 (not shown in Fig. 34 but shown in Fig. 1) such that the springs 418 15 are compressed as the pressure ring shell 412 and the pistons 402 move upwardly (in the view of Fig. 34). The optical core 420 includes a block member 421 as a separate element to permit ease of machining of optical core 420. The optical core 420 has an axially extending slot 424 20 (defined by walls 427) on a forward side thereof (aligned with slot 408 on front clam shell 406) and sized to receive the sampler 410 as will be described. Further, the pressure ring shell 412 has an axial slot 413 (aligned with slot 408). The walls 427 are received within slots 408 and 25 413.

An optics housing 430 is secured to the optical core 420 for movement therewith. The optics housing 430 carries an optical source 432 as well as optical detectors 434 and a beam splitter 436 (shown only in Fig. 34). The

optical source 432 directs infrared light to the beam splitter 436 which splits the beams into signals, each directed to detectors 434, to enhance optical analyzing of the signal. The source 432 directs the infrared light

5 through an optical path which passes through a slot 431 formed in the optics housing 430. Slot 431 is aligned with slots 424 and 408. A ball plunger 438 is urged by a spring 440 into slot 436. A cam pin 442 is carried in the deployment block 421 in order to protrude into the slot

10 424. Finally, a fan is preferably provided for drying any sample within the apparatus. While the fan impeller is not shown, the fan shroud 444 is shown into which a fan impeller may be placed for blowing air through the optical core 420 and the optics housing 430.

15 An enhanced design sampler 410 includes a handle end 409 and a sample end 411. The handle end 409 is a hollow sampler housing. The interior 500 of end 409 is sized to completely receive the sample end 411. The housing 409 is sized to be received within the slot 424.

20 As best shown in Fig. 36, side walls of the housing 409 have bottom edges 501 which are positioned against and ride on inwardly protruding rails 425 of the optical core 420 at the bottom of slot 424. The sample end 411 is pivotly secured to the sampler housing 409 at a

25 pivot pin 502. As a result, the sample end 411 can pivot between a storage position shown in Fig. 36 with the sampler end 411 fully received within the interior 500 of handle 409 in order to protect the sample end 411 and its

constituent components from destructive impact or contamination.

The sample end 411 can pivot from the storage position of Fig. 36 to a sample position shown in Fig. 39.

- 5 The sample end 411 contains the membrane 504 for receiving a sample of interstitial fluid. Further, the sample end 411 contains a hub 506 terminating at a pressure ring 508. The hub 506 contains a needle 510 for collecting interstitial fluid and depositing the interstitial fluid on 10 the membrane 501 as previously described.

In the storage position of Fig. 36, the membrane 504 is contained with the housing 409 to prevent damage as well as contamination (such as from finger prints or other skin oil). Further, in the storage position, the needle 15 510 is protected from damage as well as protecting a user from undesired contact with the needle 510.

In the sample position of Fig. 39 with the sampler 410 fully inserted, the membrane 504 is positioned in the optical pathway, T, between the light source and the 20 light detectors. Further, the needle 510 is positioned centrally aligned with the pressure ring 415. The hub 506 and ring 508 are sized to freely pass through the ring 415. Also, in the position of Fig. 39, the ball plunger 438 (shown only in Fig. 33) moves along an axis Y-Y to be 25 received within in a detent 512 formed within the sample end 411 in order to retain the sample end 411 in position.

When in the sample position, flanges 514 on the sample end 411 oppose and abut against a stop surface 516 on the optics housing 430, (Fig. 34). As a result, the

opposition of the flanges 514 and surface 516 prevents upward movement of the sampler relative to the optics housing 430. Simultaneously, opposition of the bottom edges 501 to the rails 425 prevents downward movement of  
5 the sampler 410 relative to the optics housing 430. The reception of the ball plunger 438 within the detent 512 restricts left and right movement (within the view of Fig. 39) of the sampler 410 relative to the apparatus. Further, such reception of the ball plunger 438 within the detent  
10 512 provides a tactile sensation to the user indicating that the sampler 410 has been fully seated with the apparatus. With the sampler 410 fully received with the apparatus, the membrane 504 is positioned within the light path, T, and the needle 510 is axially aligned with the  
15 hub.

The sample end 411 is pivoted to the sample position of Fig. 39 as the housing 409 is urged within the slot 424. Specifically, the cam 442 abuts a first cam follower 520 on the sample end 411. Figs. 36 through 39 illustrate the sequence of operation for pivoting the end 411 to the sample position. In Fig. 36, the sample end 411 is in the storage position and the handle 409 is in the process of initial insertion into the slot 424 with the bottom edges 501 of housing 409 riding on rails 425. The  
20 cam pin 442 is opposing the first cam follower 520. Upon further insertion of the housing such that the leading end 503 of the sample end 411 moves toward abutment with a wall 505 of the optical core 420 (Fig. 36), the cam pin 442 pushes against the cam follower 520 causing pivoting  
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movement of the sample end 411 about the pivot pin 502 as illustrated in Fig. 37. Fig. 38 illustrates full pivotal movement of end 411 with end 411 not yet fully advanced to a full seated position. Fig. 39 illustrates full insertion 5 of sampler 410 with the detent 512 aligned with the axis Y-Y of the pin 438 and with the membrane 504 aligned with the optical pathway, T. The compression of cam pin 442 against surface 520 prevents downward pivoting of end 411 and edge 514 against stop surface 516 (Fig. 34) prevents upward 10 pivoting thereby locking end 411 in place with pin 438 in detent 512 and with the target area of membrane 504 in the light path. At this point, the apparatus can be used by urging the ring end 415 against the skin (preferably in the arm region). In response to such urging, the pressure ring 15 shell 412 and attached hollow pistons 402 move upwardly to compress the springs 418. Following initial upward movement of the ring 415 relative to the hub 506, the needle penetrates the skin and the hub end 508 further engages and pushes against the skin with causation of 20 movement of the optical core 420 and attached optics housing 430 and sampler 410 against the urging of the springs 416. The thus described dual operation of two pressure rings 415, 508 operating against the urging of two springs 416, 418 is more fully described with reference to 25 Figs. 28 through 31.

After a fluid sample has been collected and/or tested, the patient can remove the tool from the arm such that the components return to the relative positioning shown in Fig. 34. At this point, the user can grasp the

handle 409 to pull the sampler 410 out of the apparatus. This pulling causes the detent 512 to become disengaged from the ball plunger 438 and further causes the cam pin 442 to engage a second cam follower 522 as illustrated in 5 Fig. 40. The engagement of the cam pin 442 with the second cam follower 522 urges the sampler end 411 to pivot from the sample position toward the storage position. An internal wall 580 acts as a stop to limit movement of end 411 and hold it in the full storage position.

10           The interior 500 of the housing contains a retention spring 524 on the housing 409. The retention spring 524 acts against an arcuate surface 526 on the sample end 411. The surface 526 terminates at a detent 528 to receive the retention spring 524. Therefore, when the 15 sample end 411 is in the storage position of Fig. 36, the retention spring 524 is received within the detent 528 preventing movement of the sample end 411 out of the storage position until an adequate force acts on the cam follower 520 to cause pivoting movement around pivot pin 20 502.

When the sample end 411 is in the sample position of Fig. 38, the spring 524 has no deflection and is at rest. However, as the sample end 411 is moved from the sample position toward to the storage position, the arcuate 25 surface 526 engages the spring 524 causing the spring 524 to deflect and create a spring force. When the pin 442 acts against the second cam surface 522 (Fig. 40) to partially move the sample end 411 to the storage position, the deflected spring 524 is received within the detent 528

to thereby urge the sample end 411 to the full storage position and retain the sample end 411 in the full storage position.

Having described this second improved embodiment,  
5 it will be noted that the needle 510 and the membrane 504 are fully protected when the sampler 410 is not inserted into the apparatus. The membrane 504 and the needle 510 are only deployed after the housing 409 is initially inserted into the apparatus (*i.e.*, into slot 424) with the  
10 cam pin 442 engaging the first cam follower 520.

### C. Additional Enhanced Embodiments

In the embodiment of Fig. 1 and as disclosed in the aforementioned Patent Application No. 08/321,305 and 15 08/136,304 corresponding to PCT International Publication No. WO95-10223, the sampling needle such as needle 10 terminates at a free end 11 which has the plane of its opening perpendicular to the plane of the membrane 12. Stated another way, the longitudinal axis of the needle 10 20 is parallel to and spaced from the surface of the membrane 12.

The present embodiment of Figs. 33-40 use an enhanced positioning of the needle 510 relative to the membrane 504. With attention to Figs. 42 and 43, the 25 sample end 411 includes a recess 600 sized to receive the needle 510. Further, the sample end includes a raised circular ring 602 surrounding the opening 604 through which the IR light is to pass. Between the ring 602 and the body of the sample end 411 an annular recess 606 is provided.

The free end 601 of the needle 510 passes through the recess 600 and nearly abuts the ring 602. Accordingly, the membrane 504 can be placed on the ring 602 and depressed into the recess 606. The membrane 504 can then be secured

5 within the recess 606 through any suitable means such as ultrasonic welding or the like. With this embodiment, the open end 601 is now disposed parallel to opposing material of the membrane 504 in a transfer area, A, of the membrane rather than perpendicular to it. Stated another way, the

10 longitudinal axis of the needle is perpendicular to opposing membrane material in the transfer area, A, with open end 601 directly abutting (or at least in close proximity to) the membrane material. As a result, fluid flowing from the needle 510 need not make a 90° bend to be

15 deposited on membrane 504. Instead, the fluid can flow directly onto the membrane 504 and be wicked on to the membrane material to a test area, T, covering opening 604. This enhances the transfer of fluid from the needle 510 to the membrane 504. Further, with this arrangement, the

20 material of the membrane 504 can act as a filter to filter out undesirable blood cells or cell fragments which may, from time to time, be transferred through the needle 510 to the membrane 504. The transfer of blood cells or cell fragments onto the membrane 504 can also be reduced by

25 providing a protein filter on the membrane in the region 610 (a filter area). Such a protein filter can be a physical filter or a filter composed of a binding agent (for example a chemical in that region to which blood, cell fragments, or other proteins may naturally attach to

prevent further transfer of the proteins into the target, T, area of the membrane). Examples of such binding agents are diethanolamine, carboxymethylcellulose, quaternary amines, and anti-RBC antibodies.

Having disclosed the present invention and a preferred embodiment, it will be appreciated that modifications and equivalents of the disclosed concepts may readily occur to one skilled in the art. It is intended that such modifications and equivalents shall be included within the scope of the claims which are appended hereto.